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(FILE 'HOME' ENTERED AT 12:29:32 ON 13 MAR 2002)

INDEX 'ADISALERTS, ADISINSIGHT, ADISNEWS, AGRICOLA, ANABSTR, AQUASCI,
BIOBUSINESS, BIOCOMMERCE, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO,
CABA,
CANCERLIT, CAPLUS, CEABA-VTB, CEN, CIN, CONFSCI, CROPB, CROPU, DDFB,
DDFU, DGENE, DRUGB, DRUGLAUNCH, DRUGMONOG2, ...' ENTERED AT 12:29:46 ON
13 MAR 2002

SEA (LACTIC(W) ACID(W) BACTERIA)

5 FILE ADISALERTS
1 FILE ADISINSIGHT
2* FILE ADISNEWS
2510 FILE AGRICOLA
16 FILE ANABSTR
106 FILE AQUASCI
1112 FILE BIOBUSINESS
76 FILE BIOCOMMERCE
3974 FILE BIOSIS
625 FILE BIOTECHABS
625 FILE BIOTECHDS
1352 FILE BIOTECHNO
5952 FILE CABA
62 FILE CANCERLIT
6663 FILE CAPLUS
446 FILE CEABA-VTB
1 FILE CEN
23 FILE CIN
82 FILE CONFSCI
1 FILE CROPB
8 FILE CROPU
16 FILE DDFB
9 FILE DDFU
938 FILE DGENE
16 FILE DRUGB
2 FILE DRUGLAUNCH
11 FILE DRUGU
38 FILE EMBAL
1203 FILE EMBASE
1228 FILE ESBIODASE
35 FILE FOMAD
24 FILE FOREGE
5135 FILE FROSTI
5585 FILE FSTA
466 FILE GENBANK
55 FILE HEALSAFE
186 FILE IFIPAT
1697 FILE JICST-EPLUS
29 FILE KOSMET
1994 FILE LIFESCI
11 FILE MEDICONF
1303 FILE MEDLINE
1 FILE NIOSHTIC
31 FILE NTIS
27 FILE OCEAN
4532 FILE PASCAL

1 FILE PHAR
1 FILE PHIC
23 FILE PHIN
325 FILE PROMT
4431 FILE SCISEARCH
1222 FILE TOXCENTER
547 FILE TOXLIT
910 FILE USPATFULL
1 FILE USPAT2
1528 FILE WPIDS
1528 FILE WPINDEX

L1 QUE (LACTIC(W) ACID(W) BACTERIA)

FILE 'CAPLUS, CABA, FROSTI, FSTA, PASCAL, BIOSIS, SCISEARCH' ENTERED AT
12:32:13 ON 13 MAR 2002

L2 441 S L1 AND (BETA-GALACTOSIDASE)
L3 0 S L2 AND (4000(W)MILLER UNITS)
L4 0 S L2 AND (MILLER UNITS)
L5 0 S L2 AND MILLER
L6 59670 S BETA-GALACTOSIDASE
L7 53 S L6 AND (MILLER(W)UNITS)
L8 0 S L7 AND (4000(W)MILLER)
L9 19 DUP REM L7 (34 DUPLICATES REMOVED)

=> d 19 ibib ab 1-19

L9 ANSWER 1 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 1
ACCESSION NUMBER: 2002:15400 CAPLUS
TITLE: Measuring **.beta.-Galactosidase**
Activity in Bacteria: Cell Growth, Permeabilization,
and Enzyme Assays in 96-Well Arrays
AUTHOR(S): Griffith, Kevin L.; Wolf, Richard E., Jr.
CORPORATE SOURCE: Department of Biological Sciences, University of
Maryland Baltimore County, Baltimore, MD, 21250, USA
SOURCE: Biochemical and Biophysical Research Communications
(2002), 290(1), 397-402
CODEN: BBRCA9; ISSN: 0006-291X
PUBLISHER: Academic Press
DOCUMENT TYPE: Journal
LANGUAGE: English
AB We describe a high-throughput procedure for measuring **.beta.-galactosidase** activity in bacteria. This procedure is unique because all manipulations, including bacterial growth and cell permeabilization, are performed in a 96-well format. Cells are permeabilized by chloroform/SDS treatment directly in the 96-well blocks and then transferred to 96-well microplates for std. colorimetric assay of **.beta.-galactosidase** activity as described by Miller [J. H. Miller (1972) Expts. in Mol. Genetics, Cold Spring Harbor Lab. Press, Cold Spring Harbor, NY]. Absorbance data are collected with a microplate reader and analyzed using a Microsoft Excel spreadsheet. The **.beta.-galactosidase** specific activity values obtained with the high-throughput procedure are identical to those obtained by the traditional single-tube method of Miller. Thus, values obtained with this procedure may be expressed as **Miller units** and compared directly to **Miller units** reported in the literature. The 96-well format for permeabilization and assay of enzyme specific activity together with the use of 12-channel and repeater pipettors enables efficient processing of hundreds of samples in an 8-h day. (c) 2002 Academic Press.
REFERENCE COUNT: 7 THERE ARE 7 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE
FORMAT

L9 ANSWER 2 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 2
ACCESSION NUMBER: 2001:170031 CAPLUS
DOCUMENT NUMBER: 135:267885
TITLE: pAM401-based shuttle vectors that enable overexpression of promoterless genes and one-step purification of tag fusion proteins directly from *Enterococcus faecalis*
AUTHOR(S): Fujimoto, Shuhei; Ike, Yasuyoshi
CORPORATE SOURCE: Department of Microbiology School of Medicine, Gunma University, Gunma, 371-8511, Japan
SOURCE: Appl. Environ. Microbiol. (2001), 67(3), 1262-1267
CODEN: AEMIDF; ISSN: 0099-2240
PUBLISHER: American Society for Microbiology
DOCUMENT TYPE: Journal
LANGUAGE: English
AB Two novel *Enterococcus faecalis*-*Escherichia coli* shuttle vectors that

utilize the promoter and ribosome binding site of bacA on the E. faecalis plasmid pPD1 were constructed. The vectors were named pMGS100 and pMGS101. PMGS100 is designed to overexpress cloned genes in E. coli and E. faecalis and encodes the bacA promoter followed by a cloning site and stop codon. PMGS101 was designed for the overexpression and purification of a cloned protein fused to a Strep-tag consisting of 9 amino acids at the carboxyl terminus. The Strep-tag provides the cloned protein with an affinity to immobilized streptavidin that facilitates protein purification.

The

authors cloned a promoterless **.beta.-galactosidase** gene from E. coli and cloned the traA gene of the E. faecalis plasmid

pAD1

into the vectors to test gene expression and protein purification., respectively. **.beta.-Galactosidase** was expressed in E. coli and E. faecalis at levels of 103 and 10 **Miller units**, respectively.

By cloning the pAD1 traA into pMGS101, the protein could be purified directly from a crude lysate of E. faecalis or E. coli with an

immobilized

streptavidin matrix by one-step affinity chromatography. The ability of TraA to bind DNA was demonstrated by the DNA-association. protein tag affinity chromatography method using lysates prepared from both E. coli and E. faecalis that overexpress TraA. The results demonstrated the usefulness of the vectors for the overexpression and cis/trans analysis of regulatory genes, purification and copurification of proteins from E. faecalis, DNA binding analysis, determination of translation initiation site, and other applications that require proteins purified from E. faecalis.

REFERENCE COUNT: 39 THERE ARE 39 CITED REFERENCES AVAILABLE FOR THIS

RECORD. ALL CITATIONS AVAILABLE IN THE RE

FORMAT

L9 ANSWER 3 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 3
ACCESSION NUMBER: 2001:167568 CAPLUS
DOCUMENT NUMBER: 134:339564
TITLE: Characterization of an oxygen-dependent inducible promoter, the nar promoter of Escherichia coli, to utilize in metabolic engineering
AUTHOR(S): Han, Se Jong; Chang, Ho Nam; Lee, Jongwon
CORPORATE SOURCE: Department of Chemical Engineering and Bioprocess Engineering Research Center, Taejeon, S. Korea
SOURCE: Biotechnology and Bioengineering (2001), 72(5), 573-576
CODEN: BIBIAU; ISSN: 0006-3592
PUBLISHER: John Wiley & Sons, Inc.
DOCUMENT TYPE: Journal
LANGUAGE: English

AB The nar promoters, whose transcription is maximally induced under microaerobic conditions in the presence of nitrate ion, were characterized

in fed-batch culture to determine whether they can be used for metabolic engineering, by which overall production of valuable chemicals can be increased. For this purpose, we tested whether the expression level of a reporter gene, the lacZ gene from the nar promoter, could be maintained constant throughout the induction period by manipulation of dissolved oxygen (DO) levels at a given nitrate ion concentration. First, E. coli was grown under aerobic conditions (DO 80%) to absorbance at 600 nm (OD600) of 35, then the nar promoter was induced by reduction of DO to different levels, combined with different frequencies and duration of alternating microaerobic and aerobic conditions throughout the entire induction period. For a wild-type nar promoter (pMW61) in a mutant host E. coli with a mutation

in

the narG gene on the chromosome of the host (RK5265), it was possible to maintain production of **.beta.-galactosidase** activity per cell (specific **.beta.-galactosidase** activity) at a constant rate at 5000, 10,000, 15,000, and 20,000 **Miller units**, using different combinations of nitrate ion concentrations. (0.1%,

0.5%, and 1%) and DO levels. In addn., it was possible to maintain prodn.

of specific **.beta.-galactosidase** activity at a constant rate at about 10,000 **Miller units** in the absence of nitrate ion when a nitrate-independent **nar** promoter (pMW618) in the **narL-** mutant of the W3110 *E. coli* strain (W3110**narL-**) was used. Based on these results, we conclude that the **nar** promoter system provides a convenient expression system for metabolic engineering as well as for maximal prodn. of recombinant proteins under conditions of fed-batch culture.

REFERENCE COUNT: 22 THERE ARE 22 CITED REFERENCES AVAILABLE FOR THIS

RECORD. ALL CITATIONS AVAILABLE IN THE RE

FORMAT

L9 ANSWER 4 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 4

ACCESSION NUMBER: 2001:6607 CAPLUS

DOCUMENT NUMBER: 134:126627

TITLE: Coupling the T7 A1 promoter to the runaway-replication

vector as an efficient method for stringent control and high-level expression of **lacZ**

AUTHOR(S): Chao, Yun-Peng; Chern, Jong-Tzer; Wen, Chih-Sheng

CORPORATE SOURCE: Department of Chemical Engineering, Feng Chia University, Taichung, Taiwan

SOURCE: Biotechnology Progress (2001), 17(1), 203-207
CODEN: BIPRET; ISSN: 8756-7938

PUBLISHER: American Chemical Society

DOCUMENT TYPE: Journal

LANGUAGE: English

AB An expression vector characterized by tight regulation and high expression

of cloned genes appears to be indispensable for the engineering need. To achieve this goal, in assocn. with **lacI** the T7 A1 promoter contg. two synthetic **lac** operators was constructed into a runaway-replication vector.

To further examine this vector system, **lacZ** was subcloned and placed under

the control of the T7 A1 promoter on the plasmid. With the application of

the thermal induction alone, the *Escherichia coli* strain harboring the recombinant plasmid was able to produce 15,000 **Miller units** of **.beta.-galactosidase**, while it yielded the recombinant protein with 45,000-50,000 **Miller units** upon both thermal and chem. induction. In sharp contrast, only 60-90 **Miller units** of **.beta.-galactosidase** was obtained for the cell at an uninduced state. As a result, the prodn. yield of **.beta.-galactosidase** over the background level is amplified approx. 170-fold by thermal induction and 500-fold by thermal and chem. induction. To produce the recombinant protein on a large scale, an approach by connecting two fermenters in series was newly developed. By applying the three-stage temp. shift in this dual fermenter system, 55,000 **Miller units** of **.beta.-galactosidase** was obtained.

Overall, it shows the potential use of the vector system developed here for its tight control and high prodn. of recombinant proteins.

REFERENCE COUNT: 22 THERE ARE 22 CITED REFERENCES AVAILABLE FOR THIS

RECORD. ALL CITATIONS AVAILABLE IN THE RE

FORMAT

L9 ANSWER 5 OF 19 FSTA COPYRIGHT 2002 IFIS

ACCESSION NUMBER: 2001(06):B0823 FSTA

TITLE: Characterization of an oxygen-dependent inducible promoter, the **nar** promoter of *Escherichia coli*, to utilize in metabolic engineering.

AUTHOR: Se Jong Han; Ho Nam Chang; Jongwon Lee

CORPORATE SOURCE: Correspondence (Reprint) address, Jongwon Lee, Dep.
of

Biochem., Sch. of Med., Catholic Univ. of
Taegu-Hyosung, Taegu 705-034, Korea. Tel.
+82-53-650-4471. Fax +82-53-621-4106. E-mail
leejw(a)cuth.cataegu.ac.kr

SOURCE: Biotechnology and Bioengineering, (2001) 72 (5)
573-576, 22 ref.
ISSN: 0006-3592

DOCUMENT TYPE: Journal

LANGUAGE: English

AB The O₂-dependent nar promoter from Escherichia coli, which can be induced by lowering the dissolved O₂ (DO) concn. in the presence of nitrate ions, was studied to investigate its feasibility for use in metabolic engineering. Expression levels of the lacZ gene, encoding **.beta.-galactosidase**, under the control of the nar promoter were examined in E. coli grown initially under aerobic conditions

(80% DO) to an OD₆₀₀ of 35, followed by reduction of the DO

level. Using the wild-type nar promoter in E. coli containing a mutation in the narG gene (encoding a subunit of nitrate reductase), specific **.beta.-galactosidase** activity could be maintained at constant levels (5000-20 000 **Miller units**) using various nitrate ion concn. Using a nitrate-independent nar promoter in a narL^{sup}- E. coli mutant, specific **.beta.-galactosidase** activity could be maintained at a constant level of approx. 10 000 **Miller units** in the absence of nitrate ions. Results showed that the nar promoter system is a suitable expression system for metabolic engineering studies and for production of recombinant proteins in fed-batch culture.

L9 ANSWER 6 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 5

ACCESSION NUMBER: 1998:301732 CAPLUS

DOCUMENT NUMBER: 129:77365

TITLE: Isolation and characterization of three Streptococcus pneumoniae transformation-specific loci by use of a lacZ reporter insertion vector

AUTHOR(S): Pestova, Ekaterina V.; Morrison, Donald A.

CORPORATE SOURCE: Laboratory for Molecular Biology, Department of Biological Sciences, University of Illinois at Chicago, Chicago, IL, USA

SOURCE: J. Bacteriol. (1998), 180(10), 2701-2710

CODEN: JOBAAAY; ISSN: 0021-9193

PUBLISHER: American Society for Microbiology

DOCUMENT TYPE: Journal

LANGUAGE: English

AB Although more than a dozen new proteins are produced when Streptococcus pneumoniae cells become competent for genetic transformation, only a few of the corresponding genes have been identified to date. To find genes responsible for the prodn. of competence-specific proteins, a random lacZ transcriptional fusion library was constructed in S. pneumoniae by using the insertional lacZ reporter vector pEVP3. Screening the library for clones with competence-specific **.beta.-galactosidase** (**.beta.-Gal**) prodn. yielded three insertion mutants with induced **.beta.-Gal** levels of about 4, 10, and 40 **Miller units**.

In all three clones, activation of the lacZ reporter correlated with competence and depended on competence-stimulating peptide. Chromosomal loci adjacent to the integrated vector were subcloned from the insertion mutants, and their nucleotide sequences were detd. Genes at two of the loci exhibited strong similarity to parts of Bacillus subtilis com operons. One locus contained open reading frames (ORFs) homologous to

the comeA and comeC genes in B. subtilis but lacked a comeB homolog. A second

locus contained four ORFs with homol. to the B. subtilis comG gene ORFs 1

to 4, but comG gene ORFs 5 to 7 were replaced in *S. pneumoniae* with an ORF encoding a protein homologous to transport ATP-binding proteins. Genes at all three loci were confirmed to be required for transformation by mutagenesis using pEVP3 for insertion duplications or an erm cassette for gene disruptions.

L9 ANSWER 7 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 6
ACCESSION NUMBER: 1998:65317 CAPLUS
DOCUMENT NUMBER: 128:214064
TITLE: Modulation of the function of the signal receptor domain of XylR, a member of a family of prokaryotic enhancer-like positive regulators
AUTHOR(S): Salto, Rafael; Delgado, Asuncion; Michan, Carmen; Marques, Silvia; Ramos, Juan L.
CORPORATE SOURCE: Department of Biochemistry, Molecular and Cellular Biology of Plants, Estacion Experimental del Zaidin, Consejo Superior de Investigaciones Cientificas, Granada, E-18008, Spain
SOURCE: J. Bacteriol. (1998), 180(3), 600-604
CODEN: JOBAAY; ISSN: 0021-9193
PUBLISHER: American Society for Microbiology
DOCUMENT TYPE: Journal
LANGUAGE: English

AB The XylR protein controls expression from the *Pseudomonas putida* TOL plasmid upper pathway operon promoter (Pu) in response to arom. effectors.

XylR-dependent stimulation of transcription from a Pu: lacZ fusion shows different induction kinetics with different effectors. With toluene, activation followed a hyperbolic curve with an apparent K of 0.95 mM and a max. **.beta.-galactosidase** activity of 2,550 Miller units. With o-nitrotoluene, in contrast, activation followed a sigmoidal curve with an apparent K of 0.55 mM and a Hill coeff. of 2.65. M-Nitrotoluene kept the XylR regulator in an inactive transcriptional form. Therefore, upon binding of an effector, the substituent on the arom. ring leads to productive or unproductive XylR forms. The different transcriptional states of the XylR regulator are substantiated by XylR mutants. XylRE172K is a mutant regulator that is able to stimulate transcription from the Pu promoter in the presence of m-nitrotoluene; however, its response to m-aminotoluene was negligible, in contrast with the wild-type regulator. These results illustrate the importance of the electrostatic interactions in effector recognition and in the stabilization of productive and unproductive forms by the regulator upon arom. binding. XylRD135N and XylRD135Q are mutant regulators that are able to stimulate transcription from Pu in the absence of effectors, whereas substitution of Glu for Asp135 in XylRD135E resulted in a mutant whose ability to recognize effectors was severely impaired. Therefore, the conformation of mutant XylRD135Q as well as XylRD135N seemed to mimic that of the wild-type regulator when effector binding occurred, whereas mutant XylRD135E seemed to be blocked in a conformation similar to that of wild-type XylR and XylRE172K upon binding to an inhibitor mol. such as m-nitrotoluene or m-aminotoluene.

L9 ANSWER 8 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 7
ACCESSION NUMBER: 1998:803860 CAPLUS
DOCUMENT NUMBER: 130:205625
TITLE: Development of a plasmid vector for overproduction of **.beta.-galactosidase** in *Escherichia coli* by using genetic components of groEx from symbiotic bacteria in *Amoeba proteus*

AUTHOR(S): Lee, Jung Eun; Ahn, Eun Young; Ahn, Tae In
 CORPORATE SOURCE: Department of Biology Education, Seoul National University, Seoul, 151-742, S. Korea
 SOURCE: J. Microbiol. Biotechnol. (1998), 8(5), 509-516
 CODEN: JOMBES; ISSN: 1017-7825
 PUBLISHER: Korean Society for Applied Microbiology
 DOCUMENT TYPE: Journal
 LANGUAGE: English
 AB A plasmid vector, pXGPRMATG-lac-Tgx, was developed for overprod. of **.beta.-galactosidase** in Escherichia coli using the genetic components of groEx, a heat-shock gene cloned from symbiotic X-bacteria in Amoeba proteus. The vector is composed of intragenic promoters P3 and P4 of groEx, the structural gene of lac operon, transcription terminator signals of lac and groEx, and ColeI and amp' of pBluescript SKII. The optimized host, E. coli DH5.alpha., transformed with the vector constitutively produced 117,310-171,961 **Miller units** of **.beta.-galactosidase** per mg protein in crude ext. The amt. of enzyme in crude ext. was 53% of total water-sol. proteins. About 43% of the enzyme could be purified to a specific activity of 322,249 **Miller units/mg** protein after two-fold purifn., using two cycles of pptn. with ammonium sulfate and one step of gel filtration. Thus, the expression system developed in this study presents a low cost and simple method for purifying overproduced **.beta.-galactosidase** in E. coli.

REFERENCE COUNT: 34 THERE ARE 34 CITED REFERENCES AVAILABLE FOR THIS

RECORD. ALL CITATIONS AVAILABLE IN THE RE

FORMAT

L9 ANSWER 9 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 8

ACCESSION NUMBER: 1998:486413 CAPLUS

DOCUMENT NUMBER: 129:198611

TITLE: Fed-batch cultivation of an oxygen-dependent inducible

promoter system, the nar promoter in escherichia coli with an inactivated nar operon

AUTHOR(S): Han, Se Jong; Chang, Ho Nam; Lee, Jongwon

CORPORATE SOURCE: Department of Chemical Engineering and Bioprocess Engineering Research Center, KAIST, Taejeon, 305-701, S. Korea

SOURCE: Biotechnol. Bioeng. (1998), 59(4), 400-406

CODEN: BIBIAU; ISSN: 0006-3592

PUBLISHER: John Wiley & Sons, Inc.

DOCUMENT TYPE: Journal

LANGUAGE: English

AB The nar promoter of Escherichia coli is maximally induced under anaerobic or microaerobic conditions in the presence of nitrate. We previously demonstrated in batch expts. that the intact nar promoter of E. coli cloned into a pBR322-based plasmid serves as a high-level expression system in a nar mutant of E. coli (J. Lee et al., 1996). In this study, we extend characterization of the nar promoter expression system to the fed-batch culture mode, which is widely used in industrial-scale fermn. From these expts., it was found that the specific **.beta.-galactosidase** activity expressed from the lacZ gene fused to the nar promoter was maximal when host cells were grown under aerobic conditions [dissolved oxygen, (DO) = 80%] to absorbance at 600 nm (OD600) = 35 before induction of the nar promoter by lowering DO to 1-2% with alternating microaerobic and aerobic conditions. Approx. 15 h after induction, the OD600 of the culture reached 135 and the specific **.beta.-galactosidase** activity increased to 40,000 **Miller units**, equiv. to approx. 35% of the total cellular proteins. The specific 3-galactosidase activity before induction

was approx. ,000 **Miller units**, giving an induction ratio of approx. 40. Based on these results, we conclude that the nar promoter provides a convenient and effective high level expression system

under conditions of fed-batch culture.

L9 ANSWER 10 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 9
ACCESSION NUMBER: 1996:661923 CAPLUS
DOCUMENT NUMBER: 125:294710
TITLE: Characterization of an oxygen-dependent inducible
promoter system, the nar promoter, and Escherichia
coli with an inactivated nar operon
AUTHOR(S): Lee, Jintae; Cho, Moo Hwan; Lee, Jongwon
CORPORATE SOURCE: Dep. Chem. Eng., Yeungnam Univ., Kyungbuk, S. Korea
SOURCE: Biotechnol. Bioeng. (1996), 52(5), 572-578
CODEN: BIBIAU; ISSN: 0006-3592

DOCUMENT TYPE: Journal

LANGUAGE: English

AB The nar promoter of Escherichia coli, which is maximally induced under anaerobic conditions in the presence of nitrate, was characterized to see whether the nar promoter cloned onto pBR322 can be used as an inducible promoter. To increase the expression level, the nar promoter was expressed in E. coli where active nitrate reductase cannot be expressed from the nar operon on the chromosome. A plasmid with the lacZ gene expressing **.beta.-galactosidase** instead of the structural genes of the nar operon was used to simplify an assay of induction of the nar promoter. The following effects were investigated

to

find optimal conditions: methods of inducing the nar promoter, optimal nitrate and molybdate concns. maximally inducing the nar promoter, the amt. of expressed **.beta.-galactosidase**, and induction ratio (specific **.beta.-galactosidase** activity after maximal induction/specific **.beta.-galactosidase** activity before induction.). The following results were obtained from

the

expts.: induction of the nar promoter was optimal when E. coli was grown in the presence of 1% nitrate at the beginning of culture; expression of

.beta.-galactosidase was not affected by molybdate; the induction ratio was maximal, approx. 300, when the overnight culture was grown in the flask for 2.5 h (OD 1.3) before being transferred to the fermentor; the amt. of **.beta.-galactosidase** per cell and per medium vol. was maximal when E. coli was grown under aerobic conditions to OD = 1.7; then the nar promoter was induced under microaerobic conditions made by lowering dissolved oxygen level (DO) to 1-2%. After approx. 6 h of induction, OD600 became 3.2 and specific **.beta.-galactosidase** activity became 36,000 Miller units, equiv. to 35% of total cellular proteins, which was confirmed from SDS-PAGE.

L9 ANSWER 11 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 10
ACCESSION NUMBER: 1996:112659 CAPLUS
DOCUMENT NUMBER: 124:167135
TITLE: Characterization of the nar promoter to use as an inducible promoter
AUTHOR(S): Lee, Jintae; Cho, Moohwan; Hong, Eock-Kee; Kim, Kwang-Soo; Lee, Jongwon
CORPORATE SOURCE: Dep. Chem. Eng., Yeungnam Univ., 705-034, S. Korea
SOURCE: Biotechnol. Lett. (1996), 18(2), 129-34
CODEN: BILED3; ISSN: 0141-5492

DOCUMENT TYPE: Journal

LANGUAGE: English

AB The nar promoter of Escherichia coli was characterized, which is maximally

induced under anaerobic conditions in the presence of nitrate. The following results are obtained; Expression of **.beta.-galactosidase** was optimal at 1% of nitrate and was not affected much by molybdate; the amt. of **.beta.-galactosidase** per unit vol. was maximal when the nar promoter was induced at OD600 = 1.7, and when anaerobic condition was made by supplying nitrogen gas. At

the optimal condition, the ratio of **.beta.-galactosidase** between before and after induction was approx. 250 **Miller units** were approx. 500. The results showed that the **tac** promoter can be used as an inducible promoter.

L9 ANSWER 12 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 11
ACCESSION NUMBER: 1996:175149 CAPLUS
DOCUMENT NUMBER: 124:222089
TITLE: High-level expression of lacZ under control of the
tac

or **trp** promoter using runaway replication vectors in
Escherichia coli
AUTHOR(S): Kidwell, John; Kolibachuk; Dennis, Douglas
CORPORATE SOURCE: Dep. Biol., James Madison Univ., Harrisonburg, VA,
22807, USA
SOURCE: Biotechnol. Bioeng. (1996), 50(1), 108-14
CODEN: BIBIAU; ISSN: 0006-3592
DOCUMENT TYPE: Journal
LANGUAGE: English

AB To det. the utility of coupling runaway replication to the expression of
cloned genes under the control of strong promoters, lacZ transcriptional
fusions to the **trp** or **tac** promoter (**P_{trp}** or **P_{tac}**) were constructed using
plasmids in which the copy no. is thermally regulated. Cells contg.
these

plasmids were able to produce **.beta.-galactosidase** to
levels between 3700 and 46,000 **Miller units** when
induced only by a temp. upshift. The addn. of the appropriate chem.
inducer, either IPTG (isopropyl-**.beta.-D-thiogalactopyranoside**) or IAA
(3-**.beta.-indoleacrylic acid**), did not significantly enhance the thermal
induction. The **P_{tac}**-controlled and **P_{trp}**-controlled lacZ induction
differed slightly in that the **P_{tac}**-controlled thermal induction exhibited
a lag of approx. 1.5 h as compared to both chem. and thermal induction,
whereas in the case of **P_{trp}**-controlled induction, an increase in **.beta.-galactosidase** expression above background occurred
at approx. the same time regardless of the means of induction. The best
vector, a **P_{trp}**-controlled lacZ fusion carried on a runaway replication
vector having a basal copy no. of 10, was able to mediate the expression
of **.beta.-galactosidase** to approx. 40,000
Miller units of **.beta.-galactosidase**
comprising 25% of the total cell protein at 17 h postinduction under
optimal conditions for protein yield. In these cells, lysis occurred as
lacZ was maximally expressed. Under noninducing conditions, the plasmids
were stable for at least 60 generations in the absence of antibiotic in
batch culture.

L9 ANSWER 13 OF 19 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
ACCESSION NUMBER: 1997:87706 BIOSIS
DOCUMENT NUMBER: PREV199799379419
TITLE: The construction and use of promoter probe vectors for
Rhodococcus sp.
AUTHOR(S): Kayser, K. J.; Yun, C.-O.; Kilbane, J. J. II
CORPORATE SOURCE: Inst. Gas Technol., 1700 S. Mt. Prospect Rd., Des Plaines,
IL 60018 USA
SOURCE: Actinomycetes, (1996) Vol. 7, No. 2, pp. 55-65.
ISSN: 0732-0574.
DOCUMENT TYPE: Article
LANGUAGE: English

AB Three promoter probe vectors have been constructed for use within
Rhodococcus sp. They are hybrid replicons capable of replicating both in
E. coli and *Rhodococcus* species due to the presence of replication
functions derived from pUC19 and the *Rhodococcus fascians* plasmid pRF29
respectively. Promoter probe vector pRCM1 contains a promoterless gene
which encodes a membrane-associated chloramphenicol efflux protein (**cmr**)
derived from *Rhodococcus fascians* plasmid pRF2, pRCAT3 contains a
promoterless chloramphenicol acetyl transferase gene (**cat**) derived from
Tn9, and pEBC26 contains a promoterless **beta-**

galactosidase gene derived from pSVB-gal. Many derivatives of pRCM1 and pRCAT3 giving inserts that regulated expression of chloramphenicol resistance in *Rhodococcus* sp. strain IGTS8 proved to be unstable in *E. coli*, frequently yielding plasmids containing deletions. This instability was found to be largely associated with these vectors; however, some inserts of *Rhodococcus* DNA increased and others alleviated this instability. Derivatives of pEBC26 were stable both in *Rhodococcus* and *E. coli* and many DNA fragments encoding *Rhodococcus* promoters were isolated. The size of these promoter-containing DNA fragments ranged from 0.15 to 3 Kb and the level of **beta-galactosidase** expression in *Rhodococcus* hosts ranged from 0.1 to 838 **Miller units**. Promoters from *Rhodococcus* were not observed to function in *E. coli*; however, the *E. coli* *rrnB* promoter was shown to function weakly in *Rhodococcus*.

L9 ANSWER 14 OF 19 FSTA COPYRIGHT 2002 IFIS

ACCESSION NUMBER: 1996(06):B0141 FSTA
 TITLE: High-level expression of *lacZ* under control of the *tac* or *trp* promoter using runaway replication vectors in *Escherichia coli*.
 AUTHOR: Kidwell, J.; Kolibachuk, D.; Dennis, D.
 CORPORATE SOURCE: Correspondence (Reprint) address, D. Dennis, Dep. of Biol., James Madison Univ., Harrisonburg, VA 22807, USA
 SOURCE: Biotechnology and Bioengineering, (1996) 50 (1) 108-114, 24 ref.
 ISSN: 0006-3592
 DOCUMENT TYPE: Journal
 LANGUAGE: English

AB The *Escherichia coli lacZ* gene, encoding **beta-galactosidase**, was placed under control of the *trp* or *tac* promoter in the runaway replication vectors pRA95 and pRA96, in which copy number is thermally regulated. Expression of *lacZ* was examined in transformed cells containing these plasmids. Increasing the temp. increased expression of the *lacZ* gene; 41.degree.C was the optimum temp. for thermal induction of gene expression. Induction of gene expression using isopropyl-**beta**-D-thiogalactopyranoside (IPTG) or 3-**beta**-indoleacrylic acid IAA did not significantly enhance thermal induction of gene expression. In thermally induced strains harbouring the *tac* promoter, a lag period of approx. 1.5 h was observed prior to **beta-galactosidase** production; no apparent lag was observed in strains possessing the *trp* promoter. Max. **beta-galactosidase** levels (up to 46 000 **Miller units**) were produced using a *trp* promoter on pRA96, having a basal copy number of 10; enzyme levels were 25% of the total cell protein 17 h after thermal induction.

L9 ANSWER 15 OF 19 FSTA COPYRIGHT 2002 IFIS

ACCESSION NUMBER: 1997(02):B0111 FSTA
 TITLE: Characterization of an oxygen-dependent inducible promoter system, the *nar* promoter, and *Escherichia coli* with an inactivated *nar* operon.
 AUTHOR: Jintae Lee; Moo Hwan Cho; Jongwon Lee
 CORPORATE SOURCE: Correspondence (Reprint) address, Jongwon Lee, Dep. of Biochem., Sch. of Med., Catholic Univ. of Taegu-Hyosung, 3056-6, Daemyung 4-Dong, Nam-Gu, Taegu 705-034, Korea. Tel. 82-53-650-4471. Fax 82-53-621-4106
 SOURCE: Biotechnology and Bioengineering, (1996) 52 (5) 572-578, 22 ref.
 ISSN: 0006-3592
 DOCUMENT TYPE: Journal
 LANGUAGE: English
 AB The *nar* promoter of *Escherichia coli*, which is optimally induced in the

presence of nitrate under anaerobic conditions, was characterized in order to ascertain its usefulness as an inducible promoter. The nar promoter was expressed in an E. coli strain having a mutant nar operon which does not express active nitrate reductase. A plasmid containing the lacZ gene, expressing **.beta.-galactosidase**, instead of the structural genes of the nar operon was used to assay induction of the nar promoter. Optimal conditions for nar induction were analysed. Results showed that induction of the nar promoter was optimal when E. coli was grown initially in the presence of 1% nitrate. Expression of the lacZ gene was not affected by molybdate ions. The amount of **.beta.-galactosidase** per cell and per medium vol. was max. when E. coli was grown under aerobic conditions to an optical density (at 600 nm) of 1.7; induction of the nar promoter was observed by lowering dissolved O₂ to microanaerobic levels (1-2%). After approx. 6 h induction, specific **.beta.-galactosidase** activity was 36 000 **Miller units**, equivalent to 35% of total cellular proteins, which was confirmed by SDS-PAGE. The specific activity of **.beta.-galactosidase** expressed from the nar promoter was comparable to that obtained using the tac and VHB promoters.

L9 ANSWER 16 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 12
ACCESSION NUMBER: 1993:54760 CAPLUS
DOCUMENT NUMBER: 118:54760
TITLE: Experimental conditions may affect reproducibility of the **.beta.-galactosidase** assay
AUTHOR(S): Giacomini, Alessio; Corich, Viviana; Ollero, Francisco
CORPORATE SOURCE: J.; Squartini, Andrea; Nuti, Marco P.
Dip. di Biotecnol. Agrarie, Univ. Stud. Padova, Padua, Italy
SOURCE: FEMS Microbiol. Lett. (1992), 100(1-3), 87-90
CODEN: FMLED7; ISSN: 0378-1097
DOCUMENT TYPE: Journal
LANGUAGE: English

AB Several exptl. conditions and parameters contributing to the detn. of **.beta.-galactosidase** activity, as proposed in the J. H. Miller (1972) assay, were studied. This assay is based on bacterial cell permeabilization followed by spectrophotometric measurement of o-nitrophenol released from its galactoside conjugate. Use of the absorbance correction factor and the nature and concn. of permeabilizing agents were taken into account as different exptl. conditions. Reaction time, culture vol., and growth stage were investigated as equation parameters. From a quant. point of view the results, in terms of **Miller units**, are markedly affected by variation in these conditions. Therefore, to ensure reproducibility it is advisable to use const. values for all the parameters.

L9 ANSWER 17 OF 19 CAPLUS COPYRIGHT 2002 ACS
ACCESSION NUMBER: 1990:546672 CAPLUS
DOCUMENT NUMBER: 113:146672
TITLE: Low copy number plasmids for regulated low-level expression of cloned genes in Escherichia coli with blue/white insert screening capability
AUTHOR(S): Lerner, Claude G.; Inouye, Masayori
CORPORATE SOURCE: Robert Wood Johnson Med. Sch., Rutgers, State Univ., Piscataway, NJ, 08854-5635, USA
SOURCE: Nucleic Acids Res. (1990), 18(15), 4631
CODEN: NARHAD; ISSN: 0305-1048
DOCUMENT TYPE: Journal
LANGUAGE: English

AB The authors constructed pCL1920 and pCL1921, a pair of low-copy-no. plasmids which contain a 580-bp BstUI fragment that carries the lac

promoter/operator, multiple cloning sites, and lacZ fragment of pUC19 cloned in place of the polylinker region in pGB2, a SC101-derived plasmid which confers spectinomycin (50 .mu.g/mL) and streptomycin (100 .mu.g/mL) resistance in E. coli. All multiple cloning sites indicated are unique except for an addnl. EcoRI site. Plasmids pCL19020 and pCL1921 contain the BstUI fragment in opposite orientations with respect to the pGB2 sequences. In the absence of inducer the pCL1920/21 vectors do not produce detectable levels of **.beta.-galactosidase** in JM105 (lacq lacZ.DELTA.M15) cells (<2 **Miller units**). In the presence of 2 mM IPTG (isopropyl-.beta.-D-thiogalactopyranoside) the **.beta.-galactosidase** levels of the pCL1920/21 [JM105] transformants rose to 11 units, whereas the pUC19 [JM105] transformants produced 470 units; a 43-fold increase. These results are consistent with the expected 40-fold difference in plasmid copy no. between pCL1920/21 (5 copies/cell) compared to that of the pUC vectors (200 copies/cell). Thus, the pCL 1920 and pCL1921 vectors allow regulated low-level expression of genes inserted downstream of the lac promoter-operator when transformed into strains contg. the lacq gene. They should also be useful for cloning genes which may be deleterious at high copy no. Since the pCL1020/21 vectors are compatible with Cole-derived plasmids, they can be used to form stable co-transformants together with pBR322 or pUC-derived plasmids. For blue/white screening of inserts, competent host cells with the lacZ.DELTA.M15 gene are used, and the transformation mixt. is plated on LB, spectinomycin plates pre-spread with 5 .mu.L of 0.2 M IPTG and 25 .mu.L 40 mg/mL 5-bromo-4-chloro-indolyl-.beta.-D-galactopyranoside per plate.

L9 ANSWER 18 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 13
 ACCESSION NUMBER: 1988:568937 CAPLUS
 DOCUMENT NUMBER: 109:168937
 TITLE: Fermentation of lactose by Zymomonas mobilis carrying a Lac+ recombinant plasmid
 AUTHOR(S): Yanase, Hideshi; Kurii, Junn; Tonomura, Kenzo
 CORPORATE SOURCE: Coll. Agric., Univ. Osaka Prefect., Osaka, 591, Japan
 SOURCE: J. Ferment. Technol. (1988), 66(4), 409-15
 CODEN: JFTED8; ISSN: 0385-6380
 DOCUMENT TYPE: Journal
 LANGUAGE: English

AB Lac+ recombinant plasmids encoding a **.beta.-galactosidase** fused protein and lactose permease of Escherichia coli were introduced into Z. mobilis. The fused protein was expressed with 450 to 5860 **Miller units** of **.beta.-galactosidase** activity, and functioned as lactase. Raffinose uptake by Z. mobilis CP4 was enhanced in the plasmid-carrying strain over the plasmid-free strain, suggesting that the lactose permease was functioning in the organism. Z. mobilis Carrying the plasmid could produce EtOH from lactose and whey, but could not grown on lactose as sole C source. Its growth was inhibited by either galactose or the galactose liberated from lactose.

L9 ANSWER 19 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 14
 ACCESSION NUMBER: 1986:201248 CAPLUS
 DOCUMENT NUMBER: 104:201248
 TITLE: Construction of a new vector for the expression of foreign genes in Zymomonas mobilis
 AUTHOR(S): Byun, M. O. K.; Kaper, J. B.; Ingram, L. O.
 CORPORATE SOURCE: Dep. Microbiol. Cell Sci., Univ. Florida, Gainesville,
 FL, 32611, USA
 SOURCE: J. Ind. Microbiol. (1986), 1(1), 9-15
 CODEN: JIMIE7

DOCUMENT TYPE:

Journal

LANGUAGE:

English

AB Broad host range plasmids are suitable as vectors to introduce antibiotic resistance genes into *Z. mobilis*. However, attempts to use these vectors to carry other genes with enteric promoters and controlling elements have resulted in limited success due to poor expression. Thus, a promoter cloning vector was constructed in a modified pBR327 and used this vector to isolate 12 promoters from *Z. mobilis* which express various levels of **.beta.-galactosidase** in *Escherichia coli*. Four of these were then subcloned into pCVD 305 for introduction into *Z. mobilis*. All expressed **.beta.-galactosidase** in *Z. mobilis* with activities of 100-1800 **Miller units**. One of these retained a BamHI site into which new genes can be readily inserted immediately downstream from the *Z. mobilis* promoter. Genetic traits carried by pCVD 305 were initially unstable but spontaneous variants were produced during subculture in which the plasmid was resistant to curing at elevated temp. One of these variants was examd. The increased stability of this variant results from an alteration in the plasmid rather than a chromosomal mutation or from chromosomal integration.

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(FILE 'HOME' ENTERED AT 13:06:14 ON 13 MAR 2002)

INDEX 'ADISALERTS, ADISINSIGHT, ADISNEWS, AGRICOLA, ANABSTR, AQUASCI,
CABA, BIOBUSINESS, BIOCOMMERCE, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO,
CANCERLIT, CAPLUS, CEABA-VTB, CEN, CIN, CONFSCI, CROPB, CROPU, DDFB,
DDFU, DGENE, DRUGB, DRUGLAUNCH, DRUGMONOG2, ...' ENTERED AT 13:06:22 ON
13 MAR 2002

SEA LACTOCOCCUS (W) LACTIS

3 FILE ADISALERTS
1 FILE ADISINSIGHT
3* FILE ADISNEWS
1417 FILE AGRICOLA
7 FILE ANABSTR
6 FILE AQUASCI
526 FILE BIOBUSINESS
12 FILE BIOCOMMERCE
2620 FILE BIOSIS
663 FILE BIOTECHABS
663 FILE BIOTECHDS
1535 FILE BIOTECHNO
4137 FILE CABA
40 FILE CANCERLIT
2854 FILE CAPLUS
321 FILE CEABA-VTB
1 FILE CEN
9 FILE CIN
41 FILE CONFSCI
12 FILE DDFU
1111 FILE DGENE
2 FILE DRUGNL
15 FILE DRUGU
2 FILE DRUGUPDATES
16 FILE EMBAL
1463 FILE EMBASE
1157 FILE ESBIODBASE
1 FILE FOREGE
1004 FILE FROSTI
1691 FILE FSTA
1211 FILE GENBANK
4 FILE HEALSAFE
72 FILE IFIPAT
262 FILE JICST-EPLUS
19 FILE KOSMET
1563 FILE LIFESCI
1839 FILE MEDLINE
1 FILE NIOSHTIC
3 FILE NTIS
1746 FILE PASCAL
1 FILE PHAR
4 FILE PHIN
39 FILE PROMT
2800 FILE SCISEARCH
630 FILE TOXCENTER
768 FILE TOXLIT

299 FILE USPATFULL

1 FILE SPAT2

159 FILE WPIDS

159 FILE WPINDEX

L1 QUE LACTOCOCCUS(W) LACTIS

FILE 'CABA, CAPLUS, SCISEARCH, BIOSIS' ENTERED AT 13:07:27 ON 13 MAR 2002

L2 84 S L1 AND PERMEAB?

L3 45 DUP REM L2 (39 DUPLICATES REMOVED)

L3 ANSWER 36 OF 45 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 17
ACCESSION NUMBER: 1992:586759 CAPLUS
DOCUMENT NUMBER: 117:186759
TITLE: Effect of the unsaturation of phospholipid acyl
chains

on leucine transport of **Lactococcus**
lactis and membrane **permeability**
AUTHOR(S): In 't Veld, Gerda; Driessen, Arnold J. M.; Konings,
Wil N.
CORPORATE SOURCE: Dep. Microbiol., Univ. Groningen, Haren, 9751 NN,
Neth.
SOURCE: Biochim. Biophys. Acta (1992), 1108(1), 31-9
CODEN: BBACAQ; ISSN: 0006-3002
DOCUMENT TYPE: Journal
LANGUAGE: English

AB The effect of the degree of unsatn. of the phospholipid acyl chains on
the

branched-chain amino acid transport system of *L. lactis* was investigated
by the use of a membrane fusion technique. Transport activity was
analyzed in hybrid membranes composed of equimolar mixts. of synthetic
unsatd. phosphatidyl ethanolamine (PE) and phosphatidylcholine (PC) in
which the no. of cis double bonds in the 18-carbon acyl chains was
varied.

The accumulation level and initial rate of both counterflow and
protonmotive-force driven transport of leucine decreased with increasing
no. of double bonds. The redn. in transport activity with increasing no.
of double bonds correlated with an increase in the passive
permeability of the membranes to leucine. The membrane fluidity
was hardly affected by the double bond content. It is concluded that the
degree of lipid acyl chain unsatn. is a minor determinant of the activity
of the branched chain amino acid transport system, but effects strongly
the passive **permeability** of the membrane.

L3 ANSWER 37 OF 45 CABA COPYRIGHT 2002 CABI DUPLICATE 18
ACCESSION NUMBER: 92:1627 CABA
DOCUMENT NUMBER: 920449928
TITLE: The bacteriocin lactococcin A specifically
increases

permeability of lactococcal cytoplasmic
membranes in a voltage-independent,
protein-mediated

manner
AUTHOR: Belkum, M. J. van; Kok, J.; Venema, G.; Holo, H.;
Nes, I. F.; Konings, W. N.; Abee, T.; Van Belkum,
M.

J.
CORPORATE SOURCE: Department of Genetics, University of Groningen,
Kerklaan 30, 9751NN Haren, Netherlands.
SOURCE: Journal of Bacteriology, (1991) Vol. 173, No. 24,
pp. 7934-7941. 36 ref.
ISSN: 0021-9193

DOCUMENT TYPE: Journal
LANGUAGE: English

AB Lactococcin A is a bacteriocin produced by **Lactococcus**
lactis. Its structural gene has recently been cloned and
sequenced. Purified lactococcin A increased the **permeability** of
the cytoplasmic membrane of *L. lactis* and dissipated the membrane
potential. A significantly higher concentration of lactococcin A was
needed to dissipate the membrane potential in an immune strain of *L.*

Lactococcus A at low concentrations (0.029 micro g/mg of protein) inhibited secondary and phosphate-bond driven transport of amino acids in sensitive cells and caused efflux of preaccumulated amino acids. Accumulation of amino acids by immune cells was not affected by this concentration of *Lactococcus* A. *Lactococcus* A also inhibited proton

motive

force-driven leucine uptake and leucine counterflow in membrane vesicles of the sensitive strain but not in membrane vesicles of the immune

strain.

These observations indicate that *Lactococcus* A makes the membrane **permeable** for leucine in the presence or absence of a proton motive force and that the immunity factor(s) is membrane linked. Membrane vesicles of *Clostridium acetobutylicum*, *Bacillus subtilis* and *Escherichia coli* were not affected by *Lactococcus* A, nor were liposomes derived from phospholipids of *L. lactis*. These results indicate that *Lactococcus* A

acts

on the cytoplasmic membrane and is very specific towards *Lactococcus*. The combined results obtained with cells, vesicles and liposomes suggest that the specificity of *Lactococcus* A may be mediated by a receptor protein associated with cytoplasmic membrane.

L3 ANSWER 38 OF 45 SCISEARCH COPYRIGHT 2002 ISI (R)

ACCESSION NUMBER: 91:688106 SCISEARCH

THE GENUINE ARTICLE: GV187

TITLE: THE BACTERIOCIN LACTOCOCCIN-A SPECIFICALLY INCREASES
PERMEABILITY OF LACTOCOCCAL CYTOPLASMIC MEMBRANES

IN A VOLTAGE-INDEPENDENT, PROTEIN-MEDIATED MANNER

AUTHOR: VANBELKUM M J (Reprint); KOK J; VENEMA G; HOLO H; NES I
F;

KONINGS W N; ABEE T

CORPORATE SOURCE: UNIV GRONINGEN, DEPT GENET, KERKLAAN 80, 9751 NN HAREN,
NETHERLANDS (Reprint); NLVF, MICROBIAL GENE TECHNOL LAB,
N-1432 AS, NORWAY; UNIV GRONINGEN, DEPT MICROBIOL, 9751

NN

HAREN, NETHERLANDS

COUNTRY OF AUTHOR: NETHERLANDS; NORWAY

SOURCE: JOURNAL OF BACTERIOLOGY, (1991) Vol. 173, No. 24, pp.
7934-7941.

DOCUMENT TYPE: Article; Journal

FILE SEGMENT: LIFE

LANGUAGE: ENGLISH

REFERENCE COUNT: 36

ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS

AB *Lactococcus* A is a bacteriocin produced by *Lactococcus*
lactis. Its structural gene has recently been cloned and sequenced
(M. J. van Belkum, B. J. Hayema, R. E. Jeeninga, J. Kok, and G. Venema,
Appl. Environ. Microbiol. 57:492-498, 1991). Purified *Lactococcus* A
increased the **permeability** of the cytoplasmic membrane of *L.*
lactis and dissipated the membrane potential. A significantly higher
concentration of *Lactococcus* A was needed to dissipate the membrane
potential in an immune strain of *L. lactis*. *Lactococcus* A at low
concentrations (0.029-mu-g/mg of protein) inhibited secondary and
phosphate-bond driven transport of amino acids in sensitive cells and
caused efflux of preaccumulated amino acids. Accumulation of amino acids
by immune cells was not affected by this concentration of *Lactococcus* A.
Lactococcus A also inhibited proton motive force-driven leucine uptake

and

leucine counterflow in membrane vesicles of the sensitive strain but not
in membrane vesicles of the immune strain. These observations indicate
that *Lactococcus* A makes the membrane **permeable** for leucine in
the presence or absence of a proton motive force and that the immunity
factor(s) is membrane linked. Membrane vesicles of *Clostridium*
acetobutylicum, *Bacillus subtilis*, and *Escherichia coli* were not affected
by *Lactococcus* A, nor were liposomes derived from phospholipids of *L.*
lactis. These results indicate that *Lactococcus* A acts on the cytoplasmic
membrane and is very specific towards *Lactococcus*. The combined results

obtained with cells, vesicles, and liposomes suggest that the specificity of lactococcin A be mediated by a receptor protein associated with the cytoplasmic membrane.

L3 ANSWER 39 OF 45 CABA COPYRIGHT 2002 CABI DUPLICATE 19
ACCESSION NUMBER: 91:11466 CABA
DOCUMENT NUMBER: 910443783
TITLE: Characterization of lactococci and lactobacilli isolated from semihard goats' cheese
AUTHOR: Requena, T.; Pelaez, C.; Desmazeaud, M. J.
CORPORATE SOURCE: Instituto del Frio (CSIC), Ciudad Universitaria, 28040 Madrid, Spain.
SOURCE: Journal of Dairy Research, (1991) Vol. 58, No. 1, pp. 137-145. 32 ref.
ISSN: 0022-0299
DOCUMENT TYPE: Journal
LANGUAGE: English

AB Several strains of *Lactococcus lactis* var. *lactis*, *Lactobacillus casei* and *Lactobacillus plantarum* isolated from traditional goat milk cheese were studied for titratable acidity, proteolysis in milk and enzymic activities. Aminopeptidase activities were measured with whole cells and cells made permeable with Triton X-100. Caseinolytic activity was investigated using PAGE. *Lc. lactis* var. *lactis* had a level of proteolytic activity in skim milk greater than that of *Lb. casei*, while this activity in *Lb. plantarum* was very low. Alanine aminopeptidase activity was almost non-existent for all strains tested, while lysine aminopeptidase activity appeared to be of fundamentally intracellular origin. Leucine aminopeptidase activity was also greater in permeable cells than in whole cells for *Lb. casei* and *Lb. plantarum*. *Lc. lactis* var. *lactis* leucine aminopeptidase activity was greater in whole cells. No significant hydrolysis of casein was found with *Lb. casei* IFPL 725 and *Lb. plantarum* IFPL 722 made permeable with Triton X-100 after 24 h incubation with whole bovine casein.

L3 ANSWER 40 OF 45 CABA COPYRIGHT 2002 CABI
ACCESSION NUMBER: 89:65434 CABA
DOCUMENT NUMBER: 890432541
TITLE: Characterization of an aminopeptidase of *Streptococcus cremoris* AM2 and an alpha-galactosidase of *Leuconostoc lactis* CNRZ 1091
Caracterisation d'une aminopeptidase chez *Streptococcus cremoris* AM2 et d'une alpha-galactosidase chez *Leuconostoc lactis* CNRZ 1091
AUTHOR: Boquien, C. Y.; Desmazeaud, M. J.; Corrieu, G.
CORPORATE SOURCE: INRA, Lab. Genie des Procédés Biotech. Agro-alimentaires, 78850 Thiverval-Grignon, France.
SOURCE: Lait, (1989) Vol. 69, No. 1, pp. 71-81. 21 ref.
DOCUMENT TYPE: Journal
LANGUAGE: French
SUMMARY LANGUAGE: English

AB The API ZYM enzyme system was used to detect enzymic activities specific to lactic acid bacteria. A total of 20 mesophilic streptococcal and 34 leuconostoc strains were tested, and 2 enzymes selected for characterization were (i) an aminopeptidase of *Streptococcus cremoris* AM2 that hydrolysed histidyl-phenylalanine- beta -naphthylamide and (ii) an alpha -galactosidase of *Leuconostoc lactis* CNRZ 1091 that hydrolysed paranitrophenol- alpha -galactose. Activity of (i) was max. in fresh cells whilst that of (ii) was max. when the cells were permeabilized with Triton X-100. Enzymes (i) and (ii) resp. had pH optimum of 7 and 6.5, and optimum temp. was 40 deg C for both enzymes. Apparent Michaelis

constant (Km) was 0.17 and 0.73 mM for (i) and (ii) resp., and Vmax was 0.6 and 90 pmol/s or 107 c.f.u. per ml.

L3 ANSWER 41 OF 45 CABA COPYRIGHT 2002 CABI
ACCESSION NUMBER: 84:85357 CABA
DOCUMENT NUMBER: 840491300
TITLE: Fermentation process improvement by membrane technology
AUTHOR: Kyung, K. H.
CORPORATE SOURCE: Michigan State Univ., East Lansing, Michigan 48824, USA.
SOURCE: Dissertation Abstracts International, B (Sciences and Engineering), (1984) Vol. 44, No. 7, pp. 2071-2072.
Order No: DA8324736.
DOCUMENT TYPE: Journal
LANGUAGE: English

AB For continuous fermentation of glucose to ethanol using *Saccharomyces cerevisiae* ATCC 4126, substrate was fed into a continuous dialysate circuit, then transferred by membrane diffusion through an intermediate dialyser into a batch fermentor circuit; product was simultaneously withdrawn from the fermentor circuit through dialyser membranes into the dialysate circuit and out in the effluent. Cells in the fermentor were essentially immobilized, and converted substrate to product by maintenance metabolism. Advantages of the system were offset by low productivity which was apparently limited by membrane permeability. A batch fermentation system utilizing a mutualistic dialysis culture of *Streptococcus lactis* and *Candida utilis* was also investigated, the 2 organisms being inoculated into separate fermentors connected by an intermediate dialyser. Lactose was fermented by *S. lactis* to lactic acid which was dialysed into the *C. utilis* culture and utilized to produce yeast biomass. Solute exchange rate across the membrane was the primary limiting factor.

L3 ANSWER 42 OF 45 CABA COPYRIGHT 2002 CABI
ACCESSION NUMBER: 84:76732 CABA
DOCUMENT NUMBER: 840491084
TITLE: Location of peptidases outside and inside the membrane of *Streptococcus cremoris*
AUTHOR: Exterkate, F. A.
CORPORATE SOURCE: Netherlands Inst. for Dairy Res., NIZO, Ede, Netherlands.
SOURCE: Applied and Environmental Microbiology, (1984) Vol. 47, No. 1, pp. 177-183. 24 ref.
ISSN: 0099-2240
DOCUMENT TYPE: Journal
LANGUAGE: English

AB Peptidase activity determinations involving native cells of *Streptococcus cremoris* and completely disrupted cell preparations, as well as experiments concerned with peptidase activity distribution among cell fractions obtained by a damage-restrictive removal of the cell wall and release of intracellular material, suggest the presence of peptidases with distinguishable locations. Alanyl, leucyl and prolyl aminopeptidase activities are most likely located in the cell wall-membrane interface, showing no detectable association with the membrane. Lysyl aminopeptidase is present not only in this location, but also as an intracellular enzyme. Endopeptidase activity and glutamate aminopeptidase activity appear to be weakly associated with the membrane. Results of experiments concerned with permeabilization of the membrane, and findings regarding an effect of the local environment of the enzymes on their pH activity profiles are evaluated and considered as being indicative of the proposed location.

The

possible implications of these findings with respect to protein utilization during growth of the organism in milk discussed.

L3 ANSWER 43 OF 45 CABA COPYRIGHT 2002 CABI

ACCESSION NUMBER: 82:16001 CABA

DOCUMENT NUMBER: 810469483

TITLE: Regulation of methyl- beta

-D-thiogalactopyranoside-

6-phosphate accumulation in Streptococcus lactis by exclusion and expulsion mechanisms

AUTHOR: Thompson, J.; Saier, M. H., Jr.

CORPORATE SOURCE: Dep. of Biol., John Muir Coll., Univ. of California at San Diego, La Jolla, California 92093, USA.

SOURCE: Journal of Bacteriology, (1981) Vol. 146, No. 3, pp.

885-894. 40 ref.

ISSN: 0021-9193

DOCUMENT TYPE: Journal

LANGUAGE: English

AB Starved cells of Streptococcus lactis ML3 (grown previously on galactose, lactose or maltose) accumulated methyl- beta -D-thiogalactopyranoside (TMG) by the lactose:phosphotransferase system. >98% of accumulated sugar was present as a phosphorylated derivative, TMG-6-phosphate (TMG-6P).

When

a phosphotransferase system sugar (glucose, mannose, 2-deoxyglucose, or lactose) was added to the medium simultaneously with TMG, the beta -galactoside was excluded from the cells. Galactose enhanced the accumulation of TMG-6P. Glucose, mannose, lactose, or maltose + arginine, when added to a suspension of TMG-6P-loaded cells of S. lactis ML3, elicited rapid expulsion of intracellular solute. The material recovered in the medium was exclusively free TMG. Expulsion of galactoside required both entry and metabolism of an appropriate sugar, and intracellular dephosphorylation of TMG-6P preceded efflux of TMG. The rate of dephosphorylation of TMG-6P by **permeabilized** cells was increased 2-3X by adenosine 5'-triphosphate but was strongly inhibited by fluoride. S. lactis ML3 (DGr) was derived from S. lactis ML3 by positive selection for resistance to 2-deoxy-D-glucose and was defective in the enzyme IIMan component of the glucose:phosphotransferase system. Neither glucose nor mannose excluded TMG from cells of S. lactis ML3 (DGr), and these 2

sugars

failed to elicit TMG expulsion from preloaded cells of the mutant strain. Accumulation of TMG-6P by S. lactis ML3 can be regulated by two independent mechanisms whose activities promote exclusion or expulsion of galactoside from the cell.

L3 ANSWER 44 OF 45 CABA COPYRIGHT 2002 CABI

ACCESSION NUMBER: 82:19154 CABA

DOCUMENT NUMBER: 820474332

TITLE: Membrane-bound peptidases in Streptococcus cremoris

AUTHOR: Exterkate, F. A.; International Dairy Federation, Symposium

CORPORATE SOURCE: Netherlands Inst. for Dairy Res. (NIZO), Ede, Netherlands.

SOURCE: Netherlands Milk and Dairy Journal, (1981) Vol. 35, No. 3/4, pp. 328-332. 12 ref.

ISSN: 0028-209X

DOCUMENT TYPE: Journal

LANGUAGE: English

AB Review of results of various experiments, relating to peptidase activity distribution among cell fractions, **permeabilization** of the membrane, and initial reaction kinetics, leads to the conclusion that Streptococcus cremoris has a functional system of specific peptidases located within the membrane and at its outside surface. This system plays a role in protein utilization during growth of the organism in milk, by making small peptides and free amino acids available for transport through

the membrane.

L3 ANSWER 45 OF 45 CABA COPYRIGHT 2002 CABI

ACCESSION NUMBER: 80:20598 CABA

DOCUMENT NUMBER: 800458595

TITLE: Effect of external factors on manifestation of antagonistic activity by lactic streptococci

AUTHOR: Zaborskikh, E. I.

SOURCE: Biologiya mikroorganizmov i ikh ispol'zovanie v narodnom khozyaistve, (1977) pp. 99-103, 208. 10 ref.

Publisher: Gosudarstvennyi Universitet imeni A. A. Zhdanova. Irkutsk

PUB. COUNTRY: USSR

DOCUMENT TYPE: Miscellaneous

LANGUAGE: Russian

AB Cultures of 2 strains of Streptococcus lactis and 2 of Str. diacetylactis showing antagonism against Escherichia coli were filtered through membrane

filters; the filtrates and hydrolysed milk as control were inoculated with

E. coli at 20 to 2×10^8 cells/ml, and growth intensity measured by nephelometer after incubation for 24 h at 30 deg C. The filtrates inhibited E. coli growth irrespective of size of inoculum. The antimicrobial compounds formed by the strains studied were stable to heating for 30 min at 65 deg C or to boiling for 20 min; they retained their activities in acid, but were inactivated under alkaline conditions; they passed through semi-permeable membranes; they were not precipitated by trichloroacetic acid, saturated $(\text{NH}_4)_2\text{SO}_4$ solution, ammonia or ethanol; and they were not extractable by diethyl ether.